Gene	Forward Primer (5' - 3')	Reverse Primer (5' - 3')
ZmBX1	CTAG <u>CTCGAG</u> CATGGCTTTCGCGCC	GACT <u>GGATCC</u> CATATGCGTACTGTA
	CAAAAC (XhoI)	GTTTTTATTCAATC (BamHI)
ZmMYB61	TCTAGAGTATGGGGAGGCCACCGTG	<u>TCTAGA</u> GACGATACAGCAACATGA
	(XbaI)	AC (XbaI)
ZmbHLH20	<u>GAATTC</u> GCACCGTAACCTGATGGAG	<u>GGATCC</u> TCGTGTTCGTCACAGCAT
	TTC (EcoRI)	GTCG (BamHI)
ZmbHLH20-	<u>GAATTC</u> GCACCGTAACCTGATGGAG	TCTAGATCGTGTTCGCAGCATGTC
eGFP	TTC (EcoRI)	GG (XbaI)
ZmbHLH76	<u>GAATTC</u> GTCGATCGAGATGAAGAGC	<u>GGATCC</u> CTCAGCTCAGACGAGGAT
	CG (EcoRI)	CG (BamHI)
ZmbHLH76-	<u>GAATTC</u> GTCGATCGAGATGAAGAGC	TCTAGACTCAGCGACGAGGATCGT
eGFP	CG (EcoRI)	C (XbaI)

Table S1 Primers used in gene cloning and vector construction

Note: The underlined sequences are the restriction enzyme digestion sites, and the corresponding enzymes are indicated in the brackets.

Primer name	Prime sequence (5' - 3')		
amiRNA-ZmBX1-I	gaTATCGAATGAACGGCCCGCTAtctctcttttgtattcc		
amiRNA-ZmBX1-II	gaTAGCGGGCCGTTCATTCGATAtcaaagagaatcaatga		
amiRNA-ZmBX1-III	gaTAACGGGCCGTTCTTTCGATTtcacaggtcgtgatatg		
amiRNA-ZmBX1-IV	gaAATCGAAAGAACGGCCCGTTAtctacatatattcct		
amiRNA-ZmbHLH20-I	gaTCTATGGTAACGATCGGTCTAtctctcttttgtattcc		
amiRNA-ZmbHLH20-II	gaTAGACCGATCGTTACCATAGAtcaaagagaatcaatga		
amiRNA-ZmbHLH20-III	gaTAAACCGATCGTTTCCATAGTtcacaggtcgtgatatg		
amiRNA-ZmbHLH20-IV	gaACTATGGAAACGATCGGTTTAtctacatatattcct		
amiRNA-ZmbHLH76-I	gaTATACTAGAGTACTACTCCGCtctctcttttgtattcc		
amiRNA-ZmbHLH76-II	gaGCGGAGTAGTACTCTAGTATAtcaaagagaatcaatga		
amiRNA-ZmbHLH76-III	gaGCAGAGTAGTACTGTAGTATTtcacaggtcgtgatatg		
amiRNA-ZmbHLH76-IV	gaAATACTACAGTACTACTCTGCtctacatatattcct		
pRS300-A	CTGCAAGGCGATTAAGTTGGGTAAC		
pRS300-B	GCGGATAACAATTTCACACAGGAA ACAG		

 Table S2 Primers used for amiRNA plasmid construction

Note: pRS300-A and pRS300-B are primers used for amplifying the amiRNA precursors and thereafter cloning into the pM999 vector by digestion with EcoRI and BamHI. The lowercase sequences match with the plasmid pRS300 sequences and the uppercase sequences are used to replace the sequences in the pRS300 vector with the target amiRNA sequences by overlapping PCR.

Benzoxazinoid	Retention time (min)	(M+H) ⁺ /Z	Mode
DHBOA-Glc	3.521	365.95	Q3 SIM(+)
DIBOA	3.563	182.05	Q3 SIM(+)
DHBOA	4.648	182.05	Q3 SIM(+)
DIBOA-Glc	4.672	344	Q3 SIM(+)
M ₂ BOA	4.868	196	Q3 SIM(+)
HMBOA-Glc	4.942	380.1	Q3 SIM(+)
DIMBOA-Glc	5.107	373.95	Q3 SIM(+)
DIMBOA	5.208	212	Q3 SIM(+)
DIM ₂ BOA	5.236	241.95	Q3 SIM(+)
MBOA	5.766	166	Q3 SIM(+)
HDM ₂ BOA-Glc	5.834	440	Q3 SIM(+)
HDMBOA-Glc	5.851	388	Q3 SIM(+)
DIM ₂ BOA-Glc	5.863	405	Q3 SIM(+)

 Table S3 HPLC-MS running parameters

Gene	Accession Number	Primer (5' - 3')
ZmIGL	GRMZM2G046191	Forward: GCCTCATAGTTCCCGACCTC
		Reverse: GAATCCTCGTGAAGCTCGTG
ZmBX1	GRMZM2G085381	Forward: CCTGCTCGGACCCCTACAT
		Reverse: GGACCCCCGCCTCTTTCAT
ZmBX2	GRMZM2G085661	Forward: GACGAGGACGACGATAAGGACTT
		Reverse: GGCCATACTCCTTCTGAAGAGACAG
ZmBX3	GRMZM2G167549	Forward: TCCACATGAAGGGCAAGGAC
		Reverse: ATCTCCATGGTCGCGAATCC
ZmBX4	GRMZM2G172491	Forward: TGCTCGCGAACCTCATCTAC
		Reverse: GAAGCGTCATCCCGAACTGA
7 m DV5	GRMZM2G063756	Forward: AGATCATGCTCGCCAACCTC
ZMBXS		Reverse: GAACGTCTCGTCCATGCTCA
7m PV6	GRMZM6G617209	Forward: TTCTTCAACACGGACGTGAG
ZmDA0		Reverse: GCCATCGAGTCCTATGGTGT
7 0.07	GRMZM2G441753	Forward: CCAGCCACGACCCCGCCAAGG
ZmDA/		Reverse: AGTAGTGTTCTCCTTCGAGGCGCCG
ZmBX8	GRMZM2G085054	Forward: CAGCTGGAGAGAGGGGGAGAT
		Reverse: TCCTCTTCCTGATGCCCTCC
Zm BY0	GRMZM2G161335	Forward: GCCAGCTGCGACGCCCCCTTCA
ZmDA9		Reverse: CGGTACGCCATGTAGTCGC
7m BV10/11	GRMZM2G311036	Forward: CTGCAACCGCTGTTTTCCTC
ZMBA10/11	GRMZM2G336824	Reverse: CCGTGGAGATATGGCTTGCT
ZmGAPDH	GRMZM2G180625	Forward: AGCAGGTCGAGCATCTTCG
		Reverse: CTGTAGCCCCACTCGTTGTC
ZmbHI H20	GRMZM2G414252	Forward: CGATATCCCCGATGCAGACC
		Reverse: CTTCTTTGGGTACCCGGTGG
ZmbHLH76	GRMZM2G112629	Forward: TTCAAGCTGCACGAGGTCAT
		Reverse: GCGAGTGGACGGTGTAGAAT
ZmMYB61	GRMZM2G108959	Forward: GTGGACCAACTACCTGAGGC
		Reverse: GGAGGTAGGAGGCTATGGCT

Table S4 Primers used for RT-qPCR analysis of gene expression



Fig. S1. Image of isolated maize protoplasts.

Isolated maize protoplasts were imaged with a microscope under a $100\times$ magnification. Scale bar = 200 $\mu m.$













(A) The HPLC chromatogram of the benzoxazinoids; (B-N) The MS profiles of the benzoxazinoids, (B) DHBOA, (C) DIBOA-Glc, (D) DHBOA-Glc, (E) HMBOA-Glc, (F) DIMBOA, (G) DIMBOA-Glc, (H) M₂BOA, (I) DIM₂BOA, (J) MBOA, (K) DIM₂BOA-Glc, (L) HDMBOA-Glc, (M) HDM₂BOA-Glc, (N) DIBOA. Y-axis indicates ionic intensity; X-axis indicates peak time.



Fig. S3. Benzoxazinoid levels in W22 and *bx2::Ds* transposon knockout mutants.

Maize W22 and *bx2::Ds* mutants were grown until two-leaves stage. Leaves were harvested and benzoxazinoids were extracted for HPLC-MS analysis. Data are means \pm SE. Asterisks indicate significant differences between WT and *bx2::Ds* (Student's t-test; n = 4; *, P \leq 0.05, **, P \leq 0.01, ***, P \leq 0.001).



Fig. S4. *ZmBX1* transcript level in maize protoplasts after overexpressing target gene. Transcript values in controls are normalized to 1. Data = means \pm SE. Asterisks indicate significant differences between control and overexpression or gene-silenced protoplasts (Student's t-test; n = 4; *, P ≤ 0.05).



Fig. S5. The contents of HDMBOA-Glc, MBOA, M₂BOA, and DIM₂BOA-Glc in protoplasts transfected with *ZmBX1* over time.

Maize protoplasts were transfected with pM999-eGFP as the control or with pM999-ZmBX1 to overexpress or silence ZmBX1 (OE-ZmBX1). The relative contents of four main benzoxazinoids, HDMBOA-Glc, MBOA, M2BOA, and DIM2BOA-Glc in control and OE-ZmBX1 maize protoplasts at different times were quantified. Asterisks indicate significant differences between control and OE-ZmBX1 protoplasts (Student's t-test; n = 5; *, P \leq 0.05).



Fig. S6. *ZmBX1* transcript level in maize protoplasts after silencing with gene-specific amiRNA.

Transcript values in controls are normalized to 1. Data = means \pm SE. Asterisks indicate significant differences between control and overexpression or gene-silenced protoplasts (Student's t-test; n = 4; *, P ≤ 0.05).



Fig. S7. *ZmMYB61* transcript level in maize protoplasts after overexpressing target gene. Transcript values in controls are normalized to 1. Data = means \pm SE. Asterisks indicate significant differences between control and overexpression or gene-silenced protoplasts (Student's t-test; n = 4; *, P ≤ 0.05).



Fig. S8. The changes of benzoxazinoid contents induced by overexpressing *ZmMYB61*. Maize protoplasts were transfected with pM999-ZmMYB61 (OE-ZmMYB61) or pM999eGFP as the control. There are no significant differences between control and ZmMYB61overexpression protoplasts (Student's t-test; n = 5).



Fig. S9. Transcription levels of target genes after overexpression or silencing.

Maize protoplasts were transfected with empty vector (control) or vectors for overexpression of *ZmbHLH20*, and *ZmbHLH76* (OE-ZmbHLH20, and OE-ZmbHLH76, respectively) or silencing *ZmbHLH20*, and *ZmbHLH76* (ami-ZmbHLH20 and amiZmbHLH76, respectively). (a) *ZmbHLH20*, and *ZmbHLH76* transcript levels in maize protoplasts after overexpression. (b) *ZmbHLH76* and *ZmbHLH20* transcript levels in maize protoplasts after silencing with gene-specific amiRNAs. Transcript values in controls are normalized to 1. Data = means \pm SE. Asterisks indicate significant differences between control and overexpression or genesilenced protoplasts (Student's t-test; n = 4; *, P \leq 0.05).



Fig. S10. Schematic representation of the predicted bHLH protein binding motifs (E-boxes) in the promoters of benzoxazinoid biosynthesis genes.

Two-kb regions upstream of the translation start sites (TSS, +1) was used for predicting the E-boxes, which are indicated by the arrow heads.