













RNAseq data for the top 10 AA-responsive genes showing relative expression levels over indicated times in presence (red) or absence of AA (blue).

yhjX

bhsA

yhcN

prpB

AGCGCACCGCAAAGTTAAGAAACCGAATATTGGGTTTAGTCTTGTTTCATAATTGT TGCAATGAAACGCGGTGAAACATTGCCTGAAACGTTAACTGAAACGCATATTTGC GGATTAGTTCATGACTTTATCTCTAACAAATTGAAATTAAACATTTAATTTTATTAAG GCAATTGTGGCACACCCCTTGCTTTGTCTTTATCAACGCAAATAACAAGTTGATAA CAAAGGATGGGCT

yhcN **\(**108)

Fig. S2

Nucleotide sequences of promoter elements for the indicated genes used in this study.



Sensors (P_{yhjX} -eGFP, P_{bhsA} -eGFP, P_{yhcN} -eGFP and P_{prpB} -eGFP) expressed in *E.coli* cells treated with 5mM acrylic acid. eGFP fluorescence was observed over 24 hours.



The *yhcN* promoter in P_{yhcN}-eGFP was truncated by removal of 108 bases at the 5' end to generate P_{yhcN} Δ 108-eGFP. Cells harbouring this construct were treated with 5mM acrylic acid (AA). Controls include untreated and 5mM lactic acid (LA) treated cells. eGFP fluorescence was measured over 24 hours.



E.coli cells stably integrated with the acrylic acid sensor shows eGFP fluorescence when exposed to acrylic acid (AA) (5mM) for 4 hours.



RAPc8 selectant #

Fig. S6

Secondary assay indicating AA reporter gene activity for indicated RAPc8 selectants. Cells were treated with acrylamide (25mM) and fluorescence measured after 2 hours. Black, grey and white bars respectively indicate values for cells expressing WT RAPc8 and inactive E142D, E142L mutants. Values highlighted in red denote fold change in mean cell fluorescence over cells expressing WT enzyme determined by subsequent FACS analysis of top 5 selectants.