**Table S2** Conditions of tests reported in the literature to determine pathogenicity of strains of *Pseudomonas syringae* to hosts in addition to the plant species from which strains were isolated.

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
| Host of isolation  | No. plant species tested | Inoculum concentration | Volume of inoculum | Inoculation technique | Re-isolation of bacteria from inoculated plant | Quantification of bacterial growth in inoculated plants | Reference |
| Hazelnut: *Corylus avellana* | 7 | 107 CFU/mL | 10 µL | Drop applied to leaf scar | yes | no | [1] |
| Hazelnut: *Corylus avellana* | 12 | Not reported | Not reported | Wounding of shoots and twigs | no | no | [2]  |
| Cherry: Prunus × yedoens, Someiyoshino | 66 | 109 CFU/mL | ‘drop’ | Drop placed on twigs and stems wounded with pin pricks | no | no | [3] |
| Mock orange: *Philadelphus coronarius* | 10 | 109 CFU/mL | Not reported | Swab inoculation of leaves with and without wounding | no | no | [4] |
| Bird of Paradise: *Strelitzia augusta* | 3 | 106 CFU/mLfor host of origin; not reported for other spp. | Not reported | Drop placed into wounded midrib for host of origin; not reported for other spp. | no | no | [5] |
| Crucifers: *Brassica olearacea* var. *botrytis* (*Bob*), *B. rapa*, *Bob* x *B. alboglabra* | 9 | 107 CFU/mL | ‘spray until run-off’ | Spray to aerial parts | Yes for original crucifer host of isolation; no for others | no | [6] |
| Crucifers: *idem* precedent case | 4 | OD600 = 0.6(≈109 CFU/mL) | 3 mL | Spray to aerial parts | yes | no | [7] |
| Tobacco: *Nicotiana* sp. | 1 (coffee) | 108 CFU/mL | Not reported | Spray to wounded leaves | yes | no | [8] |
| Cantaloupe: *Cucumis melo* | 18 | 108 CFU/mL | 50 µL | Infiltration of leaf blade | Yes in some cases | no | [9] |
| Diverse: crucifers, tomato, snapdragon, squash | 7 | 106 CFU/mL | 10 µL | Infiltration of leaf blade | no | no | [10] |
| Diverse crops and environmental reservoirs | 5 | OD600=0.001 to 0.1 depending on host (≈106 – 108 CFU/mL) | Not reported | Application to plant surface with wounding for certain hosts | yes | yes | [11, 12] |
| Diverse weeds | 1 (bean) | 108 CFU/mL (sprays)106 CFU/mL (injections) | Not reported | Spray on or inject into leaves | yes | yes | [13] |
| Kiwi | 13 | 103 to 106 CFU/mL | 10–20 µL | Injection into wounds of leaves and shoots of woody crops | yes | no | [14] |
| Myrtle: *Myrtus communis* | 6 | 108 CFU/mL | Not reported | Injection into shoots | no | no | [15] |
| Bristle oat : Avena storigosa | 12 | 108 CFU/mL | Not reported | Spray on leaves | no | no | [16] |
| Pea, soybean, snap bean: *Pisum sativum*, *Glycine max, Phaseolus vulgaris* | 50 cultivars representing 10 spp. | Not reported | Not reported | Water soaking of leaf blades or spraying of wounded leaves | no | no | [17] |
| Cucumber: *Cucumis sativus* | 28 | 106 CFU/mL | ‘spray until run-off’ | Spraying of leaves | no | no | [18] |
| Diverse: 44 plant spp. | 1 (lilac: *Syringa vulgaris*) | 107 to 108 CFU/mL | Not reported | Spraying of wounded leaves; injection of petioles | yes | no | [19] |
| Sunflower: *Helianthus annuus* | 9 | 106, 107 & 108 CFU/mL | Not reported | Low or high pressure spraying of leaves; injection into fruits | no | no | [20] |
| Diverse wild plants and cultivated kiwifruit (*Actinidia chinensis*) | 1 (Kiwifruit: *Actinidia chinensis*) | 109 CFU/mL for spray inoculation and 108 for wound inoculation | 10 mL for spraying and 10 µL for wounds | Spraying of leaves; droplet on wound to twigs | Yes in some cases | Yes in some cases | [21] |
| Diverse fruit crops and environmental reservoirs | 20 | 108 CFU/mL | 10 µL | Drops on leaves; drop on wound to petiole or stem | Yes in some cases | Yes in some cases | [22] |
| Diverse crops and environmental reservoirs | 3 | 108 CFU/mL | 50 µL | Infiltration into leaf blade | Yes in some cases | no | [23] |

**References**

1. Scortichini M, Rossi MP, Loreti S, Bosco A, Fiori M, Jackson RW, Stead DE, Aspin A, Marchesi U, Zini M *et al*: *Pseudomonas syringae* pv. *coryli*, the causal agent of bacterial twig dieback of *Corylus avellana*. *Phytopathology* 2005, 95:1316-1324.

2. Psallidas PG: *Pseudomonas syringae* pv. *avellanae* pathovar nov., the bacterium causing canker disease on *Corylus avellana*. *Plant Pathol* 1993, 42:358-363.

3. Kamiunten H, Nakao T, Oshida S: *Pseudomonas syringae* pv. *cerasicola*, pv. nov., the causal agent of bacterial gall of cherry tree. *J Gen Plant Pathol* 2000, 66:219–224.

4. Roberts SJ: Variation within *Pseudomonas syringae* pv. *philadelphi*, the cause of a leaf spot of *Philadelphus* spp. *J Appl Bacteriol* 1985, 59:283–290.

5. Polizzi G, Castello I, Parlavecchio G, Cirvilleri G: First report of bacterial blight of *Strelitzia augusta* caused by *Pseudomonas syringae* pv. *lachrymans*. *Plant Dis* 2005, 89:1010.

6. Cintas NA, Koike ST, Bull CT: A new pathovar, *Pseudomonas syringae* pv. *alisalensis* pv. nov., proposed for the causal agent of bacterial blight of broccoli and broccoli raab. . *Plant Dis* 2002, 86:992-998.

7. Bull CT, Manceau C, Lydon J, Kong H, Vinatzer BA, Fischer-Le Saux M: *Pseudomonas cannabina* pv. *cannabina* pv. nov., and *Pseudomonas cannabina* pv. *alisalensis* (Cintas Koike and Bull, 2000) comb. nov., are members of the emended species *Pseudomonas cannabina* (ex Šutič & Dowson 1959) Gardan, Shafik, Belouin, Brosch, Grimont & Grimont 1999. *Systematic and Applied Microbiology* 2010, 33(3):105-115.

8. Destéfano SAL, Rodrigues LMR, Beriam LOS, Patricio FRA, Thomaziello RA, Rodrigues-Neto J: Bacterial leaf spot of coffee caused by *Pseudomonas syringae* pv. *tabaci* in Brazil. *Plant Pathol* 2010, 59:1162-1163.

9. Morris CE, Glaux C, Latour X, Gardan L, Samson R, Pitrat M: The relationship of host range, physiology, and genotype to virulence on cantaloupe in *Pseudomonas syringae* from cantaloupe blight epidemics in France. *Phytopathology* 2000, 90:636-646.

10. Hendson M, Hildebrand DC, Schroth N: Relatedness of *Pseudomonas syringae* pv. *tomato*, *Pseudomonas syringae* pv. *maculicola*, and *Pseudomonas syringa*e pv. *antirrhini*. *J Appl Bacteriol* 1992, 73:455-464.

11. Cai R, Yan S, Liu H, Leman S, Vinatzer BA: Reconstructing host range evolution of bacterial plant pathogens using *Pseudomonas syringae* pv. *tomato* and its close relatives as a model. *Infection, Genetics and Evolution* 2011, 11(7):1738-1751.

12. Monteil CL, Cai R, Liu H, Mechan Llontop ME, Leman S, Studholme DJ, Morris CE, Vinatzer BA: Nonagricultural reservoirs contribute to emergence and evolution of *Pseudomonas syringae* crop pathogens. *New Phytol* 2013, 199 800-811.

13. Fernández-Sanz AM, Rodicio MR, González AJC: *Pseudomonas syringae* pv. *phaseolicola* isolated from weeds in bean crop fields. *Letters in Applied Microbiology* 2016, 62(4):344-348.

14. Ferrante P, Scortichini M: Redefining the global populations of *Pseudomonas syringae* pv. *actinidiae* based on pathogenic, molecular and phenotypic characteristics. *Plant Pathology* 2015, 64(1):51-62.

15. Cinelli T, Marchi G, Cimmino A, Marongiu R, Evidente A, Fiori M: Heterogeneity of *Pseudomonas savastanoi* populations infecting *Myrtus communis* in Sardinia (Italy). *Plant Pathology* 2014, 63(2):277-289.

16. Ishiyama Y, Yamagishi N, Ogiso H, Fujinaga M, Takikawa Y: Bacterial brown spot on *Avena storigosa* Schereb. caused by *Pseudomonas syringae* pv. *alisalensis*. *J Gen Plant Pathol* 2013, 79(2):155-157.

17. Hunter PJ, Taylor JD: Patterns of interaction between isolates of three pathovars of *Pseudomonas syringae* and accessions of a range of host and nonhost legume species. *Plant Pathol* 2006, 55(1):46-53.

18. Goumas DE, Chatzaki AK: Characterization and host range evaluation of *Pseudomonas viridiflava* from melon, blite, tomato, chrysanthemum and eggplant. *European Journal of Plant Pathology* 1998, 104(2):181-188.

19. Young JM: Pathogenicity and identification of the lilac pathogen, *Pseudomonas syringae* pv. *syringae* van Hall 1902. *Ann Appl Biol* 1991, 118:283-298.

20. Arsenijevic M, Vennete RJ, Masirevic S: *Pseudomonas syringae* pv. *helianthi* (Kawamura 1934) Dye, Wilke et Young 1978, a pathogen of sunflower (*Helianthus annuus* L.). *J Phytopathol* 1994, 142:199-208.

21. Liu P, Xue S, He R, Hu J, Wang X, Jia B, Gallipoli L, Mazzaglia A, Balestra GM, Zhu L: *Pseudomonas syringae* pv. *actinidiae* isolated from non-kiwifruit plant species in China. *European Journal of Plant Pathology* 2016, 145(4):743-754.

22. Bartoli C, Lamichhane JR, Berge O, Guilbaud C, Varvaro L, Balestra G, Vinatzer BA, Morris CE: A framework to gage the epidemic potential of plant pathogens in environmental reservoirs: the example of kiwifruit canker *Molec Plant Pathol* 2015, 16:137-149.

23. Morris CE, Sands DC, Vinatzer BA, Glaux C, Guilbaud C, Buffière A, Yan S, Dominguez H, Thompson BM: The life history of the plant pathogen *Pseudomonas syringae* is linked to the water cycle. *ISME Journal* 2008, 2:321-334.